

Stochastic signaling network mediates the probabilistic induction of long-term depression

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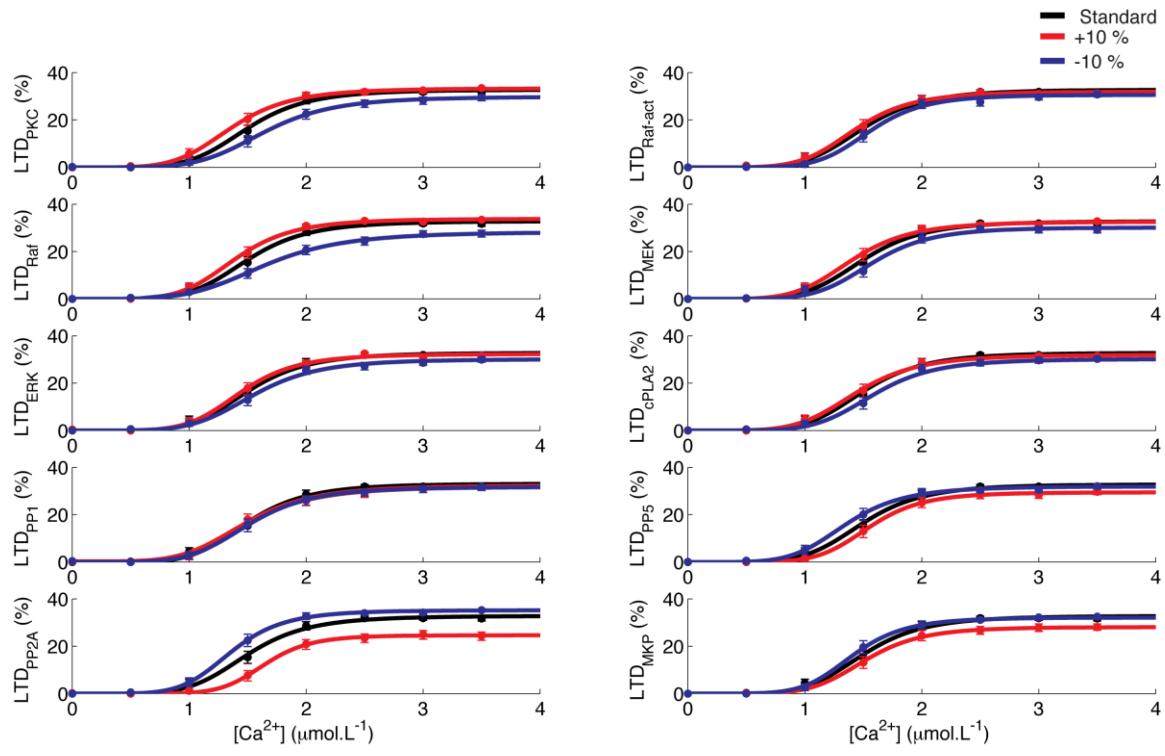


Figure 1 Sensitivity analysis of the population size of the different components of the positive feedback loop. The curve obtained with the standard model (Standard) was used for comparison. Ca^{2+} -induced LTD (pulses of 4 s and $0.0\text{-}3.5 \mu\text{mol.L}^{-1}$) was used to verify the role of small variations in the population size of each component of the feedback loop on the macroscopic behavior of the model. Data are plotted as means \pm SEM.

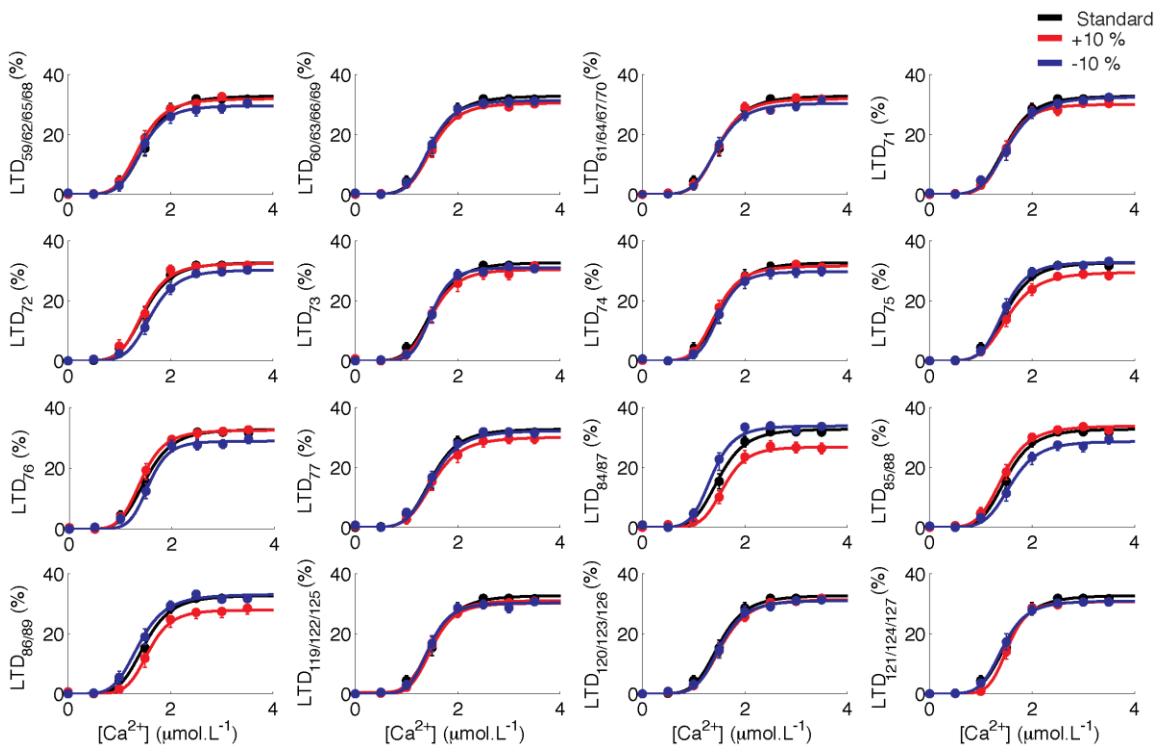


Figure 2 Sensitivity analysis of the rate constants of the positive feedback loop. The number indicated in the y-axis corresponds to the identity of the rate constant of the reactions (reac) shown in Supplementary Table I. Ca^{2+} -induced LTD (pulses of 4 s and 0.0-3.5 $\mu\text{mol.L}^{-1}$) was used to verify the role of variation in the rate constants on the macroscopic behavior of the model. The standard model was included for comparison. Data are plotted as means \pm SEM.

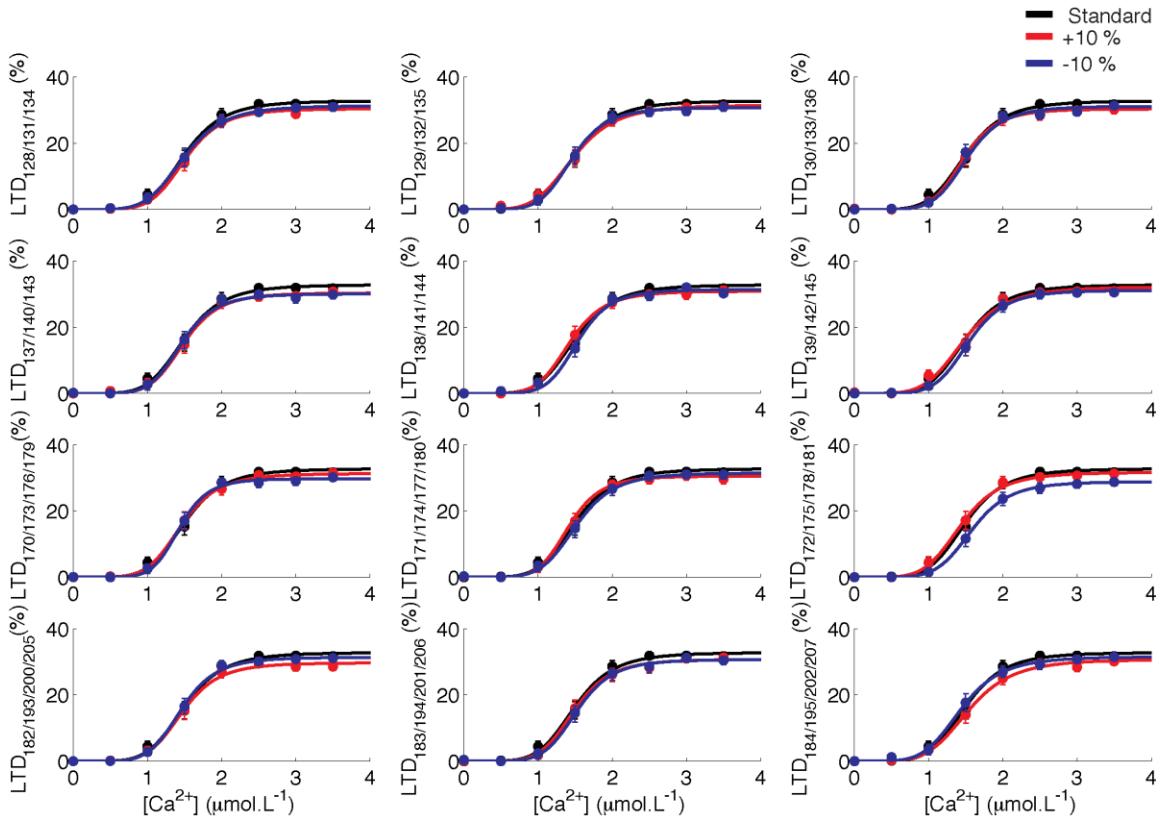


Figure 3 Continuation of the sensitivity analysis of the rate constants of the positive feedback loop. The number indicated in the y-axis corresponds to the identity of the rate constant of the reactions (reac) shown in Supplementary Table I. The figures show results of Ca^{2+} -induced LTD (pulses of 4 s and $0.0\text{-}3.5 \mu\text{mol.L}^{-1}$). Data are plotted as means \pm SEM.

Table 1 Parameters used in the model of cerebellar LTD

Parameter identity	Parameters of the model: chemical reactions, rate constants, population and geometric dimensions	References
1	Spine volume: $0.08 \mu\text{m}^3$	(Harris and Stevens, 1988)
2	SER volume: $0.017 \mu\text{m}^3$	(Harris and Stevens, 1988)
3	PSD area: $0.15 \mu\text{m}^2$	(Harris and Stevens, 1988)
4	Basal $[\text{Ca}^{2+}] = 45 \text{ nmol.L}^{-1}$ ^a	(Schmidt et al., 2003)
5	PMA = 10 molecules	(Doi et al., 2005)
6	NCX = 3 molecules	(Doi et al., 2005)
7	SERCA = 100 molecules	(Doi et al., 2005)
8	$[\text{PV}] = 40 \mu\text{mol.L}^{-1}$	(Schmidt et al., 2003)
9	$[\text{CB}] = 40 \mu\text{mol.L}^{-1}$	(Schmidt et al., 2003)
10	PKC = 48 molecules	(Cheng et al., 2006)
11	$[\text{Raf_act}] = 0.5 \mu\text{mol.L}^{-1}$	This paper
12	$[\text{Raf}] = 0.1 \mu\text{mol.L}^{-1}$	(Huang and Ferrell, 1996; Pearson et al., 2001)
13	$[\text{MEK}] = 1.5 \mu\text{mol.L}^{-1}$	(Huang and Ferrell, 1996; Pearson et al., 2001; Fujioka et al., 2006)
14	$[\text{PP5}] = 1.0 \mu\text{mol.L}^{-1}$	(Bahl et al., 2001; Rossie et al., 2006)
15	$[\text{PP2A}] = 1.5 \mu\text{mol.L}^{-1}$	(Cheng et al., 2006)

16	$[ERK] = 1.0 \mu\text{mol.L}^{-1}$	(Huang and Ferrell, 1996; Pearson et al., 2001; Cheng et al., 2006; Fujioka et al., 2006)
17	$[MKP] = 0.26 \mu\text{mol.L}^{-1}$	(Huang and Ferrell, 1996)
18	$[cPLA_2] = 0.4 \mu\text{mol.L}^{-1}$	(Tanaka et al., 2007)
19	PP1 = 30 molecules	(Cheng et al., 2006)
20	Synaptic AMPAR ~ 120-130 molecules	(Momiyama et al., 2003; Masugi-Tokita et al., 2007)
21	GRIP = 141 molecules	This paper
Reac1	$PMCA + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})PMCA$, ^b $k_f = 25000.0 \mu\text{mol}^{-1}.\text{L.s}^{-1}$	(Doi et al., 2005)
Reac2	$(Ca^{2+})PMCA \xrightarrow{k_b} PMCA + Ca^{2+}$, $k_b = 2000.0 \text{ s}^{-1}$	(Doi et al., 2005)
Reac3	$(Ca^{2+})PMCA \xrightarrow{k_{cat}} PMCA$, $k_{cat} = 500.0 \text{ s}^{-1}$	(Doi et al., 2005)
Reac4	$NCX + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})NCX$, $k_f = 93.827 \mu\text{mol}^{-1}.\text{L.s}^{-1}$	(Doi et al., 2005)
Reac5	$(Ca^{2+})NCX \xrightarrow{k_b} NCX + Ca^{2+}$, $k_b = 612.6 \text{ s}^{-1}$	(Doi et al., 2005)
Reac6	$(Ca^{2+})NCX + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2NCX$, $k_f = 93.827 \mu\text{mol}^{-1}.\text{L.s}^{-1}$	(Doi et al., 2005)
Reac7	$(Ca^{2+})_2NCX \xrightarrow{k_b} (Ca^{2+})NCX + Ca^{2+}$, $k_b = 612.6 \text{ s}^{-1}$	(Doi et al., 2005)
Reac8	$(Ca^{2+})_2NCX \xrightarrow{k_{cat}} NCX$, $k_{cat} = 1000.0 \text{ s}^{-1}$, non-conservative reaction	(Doi et al., 2005)
Reac9	$SERCA + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})SERCA$, $k_f = 17147.0 \mu\text{mol}^{-1}.\text{L.s}^{-1}$	(Doi et al., 2005)

Reac10	$(Ca^{2+})SERCA \xrightarrow{k_b} SERCA + Ca^{2+}$, $k_b = 8426.3 \text{ s}^{-1}$	(Doi et al., 2005)
Reac11	$(Ca^{2+})SERCA + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2SERCA$, $k_f = 17147.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Doi et al., 2005)
Reac12	$(Ca^{2+})_2SERCA \xrightarrow{k_b} (Ca^{2+})SERCA + Ca^{2+}$, $k_b = 8426.3 \text{ s}^{-1}$	(Doi et al., 2005)
Reac13	$(Ca^{2+})_2SERCA \xrightarrow{k_{cat}} SERCA + 2Ca_{SER}^{2+}$, $k_{cat} = 250.0 \text{ s}^{-1}$	(Doi et al., 2005)
Reac14	$\xrightarrow{k_{leak}} Ca^{2+}$, $k_{leak} = 1900.0 \mu\text{mol} \cdot \text{L}^{-1} \cdot \text{s}^{-1}$	This paper
Reac15	$PV + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})PV$, $k_f = 107.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Schmidt et al., 2003)
Reac16	$(Ca^{2+})PV \xrightarrow{k_b} PV + Ca^{2+}$, $k_b = 0.95 \text{ s}^{-1}$	(Schmidt et al., 2003)
Reac17	$(Ca^{2+})PV + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2PV$, $k_f = 107.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Schmidt et al., 2003)
Reac18	$(Ca^{2+})_2PV \xrightarrow{k_b} (Ca^{2+})PV + Ca^{2+}$, $k_b = 0.95 \text{ s}^{-1}$	(Schmidt et al., 2003)
Reac19	$PV \xrightarrow{[Mg^{2+}]k_f} (Mg^{2+})PV$, $k_f = 472.0 \text{ s}^{-1}$	(Schmidt et al., 2003)
Reac20	$(Mg^{2+})PV \xrightarrow{k_b} PV$, $k_b = 25.0 \text{ s}^{-1}$	(Schmidt et al., 2003)
Reac21	$(Mg^{2+})PV \xrightarrow{[Mg^{2+}]k_f} (Mg^{2+})_2PV$, $k_f = 472.0 \text{ s}^{-1}$	(Schmidt et al., 2003)
Reac22	$(Mg^{2+})_2PV \xrightarrow{k_b} (Mg^{2+})PV$, $k_b = 25.0 \text{ s}^{-1}$	(Schmidt et al., 2003)
Reac23	$CB + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})CB$, $k_f = 5.5 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$, high affinity site	(Schmidt et al., 2003)
Reac24	$(Ca^{2+})CB \xrightarrow{k_b} CB + Ca^{2+}$, $k_b = 2.6 \text{ s}^{-1}$, high affinity site	(Schmidt et al., 2003)
Reac25	$(Ca^{2+})CB + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2CB$, $k_f = 5.5 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$, high affinity site	(Schmidt et al., 2003)
Reac26	$(Ca^{2+})_2CB \xrightarrow{k_b} (Ca^{2+})CB + Ca^{2+}$, $k_b = 2.6 \text{ s}^{-1}$, high affinity site	(Schmidt et al., 2003)

Reac27	$CB + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})CB,$ $k_f = 43.5 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$, medium affinity site	(Schmidt et al., 2003)
Reac28	$(Ca^{2+})CB \xrightarrow{k_b} CB + Ca^{2+},$ $k_b = 35.8 \text{ s}^{-1}$, medium affinity site	(Schmidt et al., 2003)
Reac29	$(Ca^{2+})CB + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2CB,$ $k_f = 43.5 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$, medium affinity site	(Schmidt et al., 2003)
Reac30	$(Ca^{2+})_2CB \xrightarrow{k_b} (Ca^{2+})CB + Ca^{2+},$ $k_b = 35.8 \text{ s}^{-1}$, medium affinity site	(Schmidt et al., 2003)
Reac31	$PKC + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})PKC,$ $k_f = 13.3 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac32	$(Ca^{2+})PKC \xrightarrow{k_b} PKC + Ca^{2+},$ $k_b = 12.0 \text{ s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac33	$(Ca^{2+})PKC + 2Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_3PKC,$ $k_f = 1.0 \mu\text{mol}^{-2} \cdot \text{L} \cdot \text{s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac34	$(Ca^{2+})_3PKC \xrightarrow{k_b} (Ca^{2+})PKC + 2Ca^{2+},$ $k_b = 12.0 \text{ s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac35	$(Ca^{2+})_3PKC \xrightarrow{k_f} (Ca^{2+})_3PKC^*, ^c$ $k_f = 11.3 \text{ s}^{-1}$	(Bittova et al., 2001; Newton, 2001, 2009)

Reac36	$(Ca^{2+})_3PKC^* \xrightarrow{k_b} (Ca^{2+})_3PKC,$ $k_b = 0.23 \text{ s}^{-1}$	(Bittova et al., 2001; Newton, 2001, 2009)
Reac37	$PKC + AA \xrightarrow{k_f} (AA)PKC,$ $k_f = 1.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac38	$(AA)PKC \xrightarrow{k_b} PKC + AA,$ $k_b = 10.0 \text{ s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac39	$(AA)PKC \xrightarrow{k_f} (AA)PKC^*,$ $k_f = 0.017 \text{ s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac40	$(AA)PKC^* \xrightarrow{k_b} (AA)PKC,$ $k_b = 0.0055 \text{ s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac41	$(Ca^{2+})PKC + AA \xrightarrow{k_f} (AA)(Ca^{2+})PKC,$ $k_f = 1.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1},$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac42	$(AA)(Ca^{2+})PKC \xrightarrow{k_b} (Ca^{2+})PKC + AA,$ $k_b = 10.0 \text{ s}^{-1}$	(Shinomura et al., 1991;

		Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac43	$(AA)(Ca^{2+})PKC \xrightarrow{k_f} (AA)(Ca^{2+})PKC^*,$ $k_f = 0.017 \text{ s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac44	$(AA)(Ca^{2+})PKC^* \xrightarrow{k_b} (AA)(Ca^{2+})PKC,$ $k_b = 0.0055 \text{ s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac45	$(AA)(Ca^{2+})PKC^* + 2Ca^{2+} \xrightarrow{k_f} (AA)(Ca^{2+})_3PKC^*,$ $k_f = 1.0 \mu\text{mol}^{-2} \cdot \text{L} \cdot \text{s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac46	$(AA)(Ca^{2+})_3PKC^* \xrightarrow{k_b} (AA)(Ca^{2+})PKC^* + 2Ca^{2+},$ $k_b = 12.0 \text{ s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac47	$(AA)PKC + Ca^{2+} \xrightarrow{k_f} (AA)(Ca^{2+})PKC,$ $k_f = 13.3 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)

Reac48	$(AA)(Ca^{2+})PKC \xrightarrow{k_b} (AA)PKC + Ca^{2+}$, $k_b = 12.0 \text{ s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac49	$(AA)(Ca^{2+})PKC + 2Ca^{2+} \xrightarrow{k_f} (AA)(Ca^{2+})_3PKC$, $k_f = 1.0 \mu\text{mol}^{-2} \cdot \text{L} \cdot \text{s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac50	$(AA)(Ca^{2+})_3PKC \xrightarrow{k_b} (AA)(Ca^{2+})PKC + 2Ca^{2+}$, $k_b = 12.0 \text{ s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac51	$(AA)(Ca^{2+})_3PKC \xrightarrow{k_f} (AA)(Ca^{2+})_3PKC^*$, $k_f = 11.3 \text{ s}^{-1}$	(Bittova et al., 2001; Newton, 2001, 2009)
Reac52	$(AA)(Ca^{2+})_3PKC^* \xrightarrow{k_b} (AA)(Ca^{2+})_3PKC$, $k_b = 0.23 \text{ s}^{-1}$	(Bittova et al., 2001; Newton, 2001, 2009)
Reac53	$(Ca^{2+})_3PKC + AA \xrightarrow{k_f} (AA)(Ca^{2+})_3PKC$, $k_f = 1.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$,	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac54	$(AA)(Ca^{2+})_3PKC \xrightarrow{k_b} (Ca^{2+})_3PKC + AA$, $k_b = 10.0 \text{ s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993;

		O'Flaherty et al., 2001)
Reac55	$(AA)PKC^* + Ca^{2+} \xrightarrow{k_f} (AA)(Ca^{2+})PKC^*$, $k_f = 13.3 \mu\text{mol}^{-1}\text{L.s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac56	$(AA)(Ca^{2+})PKC^* \xrightarrow{k_b} (AA)PKC^* + Ca^{2+}$, $k_b = 12.0 \text{ s}^{-1}$	(Keranen and Newton, 1997; Kohout et al., 2002; Torrecillas et al., 2004)
Reac57	$(Ca^{2+})_3PKC^* + AA \xrightarrow{k_f} (AA)(Ca^{2+})_3PKC^*$, $k_f = 1.0 \mu\text{mol}^{-1}\text{L.s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac58	$(AA)(Ca^{2+})_3PKC^* \xrightarrow{k_b} (Ca^{2+})_3PKC^* + AA$, $k_b = 10.0 \text{ s}^{-1}$	(Shinomura et al., 1991; Schaechter and Benowitz, 1993; O'Flaherty et al., 2001)
Reac59	$(Ca^{2+})_3PKC^* + Raf_act \xrightarrow{k_f} ((Ca^{2+})_3PKC^*)Raf_act$, $k_f = 5.8 \mu\text{mol}^{-1}\text{L.s}^{-1}$	(Woodgett et al., 1986)
Reac60	$((Ca^{2+})_3PKC^*)Raf_act \xrightarrow{k_b} (Ca^{2+})_3PKC^* + Raf_act$, $k_b = 3.608 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac61	$((Ca^{2+})_3PKC^*)Raf_act \xrightarrow{k_{cat}} (Ca^{2+})_3PKC^* + Raf_act^*$, $k_{cat} = 4.7 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac62	$(AA)PKC^* + Raf_act \xrightarrow{k_f} (AA)(PKC^*)Raf_act$, $k_f = 5.8 \mu\text{mol}^{-1}\text{L.s}^{-1}$	(Woodgett et al., 1986)

Reac63	$((AA)PKC^*)Raf_act \xrightarrow{k_b} (AA)PKC^* + Raf_act,$ $k_b = 3.608 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac64	$(AA)(PKC^*)Raf_act \xrightarrow{k_{cat}} (AA)PKC^* + Raf_act^*,$ $k_{cat} = 4.7 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac65	$(AA)(Ca^{2+})PKC^* + Raf_act \xrightarrow{k_f} ((AA)(Ca^{2+})PKC^*)Raf_act,$ $k_f = 5.8 \mu\text{mol}^{-1} \text{ L s}^{-1}$	(Woodgett et al., 1986)
Reac66	$((AA)(Ca^{2+})PKC^*)Raf_act \xrightarrow{k_b} (AA)(Ca^{2+})PKC^* + Raf_act,$ $k_b = 3.608 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac67	$((AA)(Ca^{2+})PKC^*)Raf_act \xrightarrow{k_{cat}} (AA)(Ca^{2+})PKC^* + Raf_act^*,$ $k_{cat} = 4.7 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac68	$(AA)(Ca^{2+})_3PKC^* + Raf_act \xrightarrow{k_f} ((AA)(Ca^{2+})_3PKC^*)Raf_act,$ $k_f = 5.8 \mu\text{mol}^{-1} \text{ L s}^{-1}$	(Woodgett et al., 1986)
Reac69	$((AA)(Ca^{2+})_3PKC^*)Raf_act \xrightarrow{k_b} (AA)(Ca^{2+})_3PKC^* + Raf_act,$ $k_b = 3.608 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac70	$((AA)(Ca^{2+})_3PKC^*)Raf_act \xrightarrow{k_{cat}} (AA)(Ca^{2+})_3PKC^* + Raf_act^*,$ $k_{cat} = 4.7 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac71	$Raf_act^* \xrightarrow{k_f} Raf_act,$ $k_b = 1.0 \text{ s}^{-1}$	(Lenzen et al., 1998; Brinkmann et al., 2002)
Reac72	$Raf_act^* + Raf \xrightarrow{k_f} (Raf_act^*)Raf,$ $k_f = 1.0 \mu\text{mol}^{-1} \cdot \text{L.s}^{-1}$	(Aksan and Kurnaz, 2003; Kiyatkin et al., 2006)
Reac73	$(Raf_act^*)Raf \xrightarrow{k_b} Raf_act^* + Raf,$ $k_b = 2.0 \text{ s}^{-1}$	(Aksan and Kurnaz, 2003; Kiyatkin et al., 2006)
Reac74	$(Raf_act^*)Raf \xrightarrow{k_{cat}} Raf_act^* + Raf^*,$ $k_{cat} = 1.5 \text{ s}^{-1}$	(Aksan and Kurnaz, 2003; Kiyatkin et al., 2006)
Reac75	$PP5 + Raf^* \xrightarrow{k_f} (PP5)Raf^*,$ $k_f = 0.55 \mu\text{mol}^{-1} \cdot \text{L.s}^{-1}$	(Aksan and Kurnaz, 2003; Kiyatkin et

		al., 2006)
Reac76	$(PP5)Raf^* \xrightarrow{k_b} PP5 + Raf^*$, $k_b = 2.0 \text{ s}^{-1}$	(Aksan and Kurnaz, 2003; Kiyatkin et al., 2006)
Reac77	$(PP5)Raf^* \xrightarrow{k_{cat}} PP5 + Raf$, $k_{cat} = 0.5 \text{ s}^{-1}$	(Aksan and Kurnaz, 2003; Kiyatkin et al., 2006)
Reac78	$Raf^* + MEK \xrightarrow{k_f} (Raf^*)MEK$, $k_f = 0.65 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Fujioka et al., 2006)
Reac79	$(Raf^*)MEK \xrightarrow{k_b} Raf^* + MEK$, $k_b = 0.065 \text{ s}^{-1}$	(Fujioka et al., 2006)
Reac80	$(Raf^*)MEK \xrightarrow{k_{cat}} Raf^* + MEK^P$, $k_{cat} = 1.0 \text{ s}^{-1}$	(Fujioka et al., 2006)
Reac81	$Raf^* + MEK^P \xrightarrow{k_f} (Raf^*)MEK^P$, $k_f = 0.65 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Fujioka et al., 2006)
Reac82	$(Raf^*)MEK^P \xrightarrow{k_b} Raf^* + MEK^P$, $k_b = 0.065 \text{ s}^{-1}$	(Fujioka et al., 2006)
Reac83	$(Raf^*)MEK^P \xrightarrow{k_{cat}} Raf^* + MEK^*$, $k_{cat} = 1.0 \text{ s}^{-1}$	(Fujioka et al., 2006)
Reac84	$PP2A + MEK^* \xrightarrow{k_f} (PP2A)MEK^*$, $k_f = 0.75 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Huang and Ferrell, 1996; Fujioka et al., 2006; Kiyatkin et al., 2006)
Reac85	$(PP2A)MEK^* \xrightarrow{k_b} PP2A + MEK^*$, $k_b = 2.0 \text{ s}^{-1}$	(Huang and Ferrell, 1996; Fujioka et al., 2006; Kiyatkin et al., 2006)
Reac86	$(PP2A)MEK^* \xrightarrow{k_{cat}} PP2A + MEK^P$, $k_{cat} = 0.5 \text{ s}^{-1}$	(Huang and Ferrell, 1996; Fujioka et

		al., 2006; Kiyatkin et al., 2006)
Reac87	$PP2A + MEK^P \xrightarrow{k_f} (PP2A)MEK^P,$ $k_f = 0.75 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Huang and Ferrell, 1996; Fujioka et al., 2006; Kiyatkin et al., 2006)
Reac88	$(PP2A)MEK^P \xrightarrow{k_b} PP2A + MEK^P,$ $k_b = 2.0 \text{ s}^{-1}$	(Huang and Ferrell, 1996; Fujioka et al., 2006; Kiyatkin et al., 2006)
Reac89	$(PP2A)MEK^P \xrightarrow{k_{cat}} PP2A + MEK,$ $k_{cat} = 0.5 \text{ s}^{-1}$	(Huang and Ferrell, 1996; Fujioka et al., 2006; Kiyatkin et al., 2006)
Reac90	$MEK^* + ERK \xrightarrow{k_f} (MEK^*)ERK,$ $k_f = 16.2 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Fujioka et al., 2006)
Reac91	$(MEK^*)ERK \xrightarrow{k_b} MEK^* + ERK,$ $k_b = 0.6 \text{ s}^{-1}$	(Fujioka et al., 2006)
Reac92	$(MEK^*)ERK \xrightarrow{k_{cat}} MEK^* + ERK^P,$ $k_{cat} = 0.15 \text{ s}^{-1}$	(Fujioka et al., 2006)
Reac93	$MEK^* + ERK^P \xrightarrow{k_f} (MEK^*)ERK^P,$ $k_f = 16.2 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Fujioka et al., 2006)
Reac94	$(MEK^*)ERK^P \xrightarrow{k_b} MEK^* + ERK^P,$ $k_b = 0.6 \text{ s}^{-1}$	(Fujioka et al., 2006)
Reac95	$(MEK^*)ERK^P \xrightarrow{k_{cat}} MEK^* + ERK^*,$ $k_{cat} = 0.3 \text{ s}^{-1}$	(Markevich et al., 2004; Fujioka et al., 2006)
Reac96	$MKP + ERK^* \xrightarrow{k_f} (MKP)ERK^*,$ $k_f = 28.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Zhao and Zhang, 2001; Zhou et al., 2002)

Reac97	$(MKP)ERK^* \xrightarrow{k_b} MKP + ERK^*$, $k_b = 0.56 \text{ s}^{-1}$	(Zhao and Zhang, 2001; Zhou et al., 2002)
Reac98	$(MKP)ERK^* \xrightarrow{k_{cat}} MKP + ERK^P$, $k_{cat} = 0.14 \text{ s}^{-1}$	(Zhao and Zhang, 2001; Zhou et al., 2002)
Reac99	$MKP + ERK^P \xrightarrow{k_f} (MKP)ERK^P$, $k_f = 13.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Zhao and Zhang, 2001; Zhou et al., 2002)
Reac100	$(MKP)ERK^P \xrightarrow{k_b} MKP + ERK^P$, $k_b = 0.396 \text{ s}^{-1}$	(Zhao and Zhang, 2001; Zhou et al., 2002)
Reac101	$(MKP)ERK^P \xrightarrow{k_{cat}} MKP + ERK$, $k_{cat} = 0.099 \text{ s}^{-1}$	(Zhao and Zhang, 2001; Zhou et al., 2002)
Reac102	$cPLA_2 + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})cPLA_2$, $k_f = 1.93 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac103	$(Ca^{2+})cPLA_2 \xrightarrow{k_b} cPLA_2 + Ca^{2+}$, $k_b = 108.0 \text{ s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac104	$(Ca^{2+})cPLA_2 + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2cPLA_2$, $k_f = 10.8 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke,

		2002)
Reac105	$(Ca^{2+})_2 cPLA_2 \xrightarrow{k_b} (Ca^{2+})_2 cPLA_2 + Ca^{2+}$, $k_b = 108.0 \text{ s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac106	$(Ca^{2+})_2 cPLA_2 \xrightarrow{k_f} (Ca^{2+})_2 cPLA_2^*$, $k_f = 300.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Stahelin et al., 2007)
Reac107	$(Ca^{2+})_2 cPLA_2^* \xrightarrow{k_b} (Ca^{2+})_2 cPLA_2$, $k_b = 15.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Stahelin et al., 2007)
Reac108	$(Ca^{2+}) cPLA_2 \xrightarrow{k_f} (Ca^{2+}) cPLA_2 memb$, $k_f = 30.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Stahelin et al., 2007)
Reac109	$(Ca^{2+}) cPLA_2 memb \xrightarrow{k_b} (Ca^{2+}) cPLA_2$, $k_b = 15.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Stahelin et al., 2007)
Reac110	$cPLA_2 \xrightarrow{k_f} cPLA_2 memb$, $k_f = 3.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002;

		Frazier et al., 2002; Stahelin et al., 2007)
Reac111	$cPLA_2memb \xrightarrow{k_b} cPLA_2,$ $k_b = 15.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Stahelin et al., 2007)
Reac112	$cPLA_2memb + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})cPLA_2memb,$ $k_f = 1.93 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac113	$(Ca^{2+})cPLA_2memb \xrightarrow{k_b} cPLA_2memb + Ca^{2+},$ $k_b = 0.41 \text{ s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac114	$(Ca^{2+})cPLA_2memb + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2cPLA_2^*,$ $k_f = 10.8 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac115	$(Ca^{2+})_2cPLA_2^* \xrightarrow{k_b} (Ca^{2+})cPLA_2memb + Ca^{2+},$ $k_b = 2.5 \text{ s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski

		and Falke, 2002)
Reac116	$(Ca^{2+})_2cPLA_2^* \xrightarrow{k_f} ((Ca^{2+})_2cPLA_2^*)APC,$ $k_f = 43.0 \text{ s}^{-1}$, pseudo-first order reaction of interaction between cPLA ₂ and its substrate (APC)	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac117	$((Ca^{2+})_2cPLA_2^*)APC \xrightarrow{k_b} (Ca^{2+})_2cPLA_2^*,$ $k_b = 600.0 \text{ s}^{-1}$	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac118	$((Ca^{2+})_2cPLA_2^*)APC \xrightarrow{k_{cat}} (Ca^{2+})_2cPLA_2^* + AA,$ $k_{cat} = 450.0 \text{ s}^{-1}$	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac119	$ERK^* + cPLA_2 \xrightarrow{k_f} (ERK^*)cPLA_2,$ $k_f = 4.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Waas and Dalby, 2002; Waas et al., 2003)
Reac120	$(ERK^*)cPLA_2 \xrightarrow{k_b} ERK^* + cPLA_2,$ $k_b = 1.0 \text{ s}^{-1}$	(Waas and Dalby, 2002; Waas et al., 2003)
Reac121	$(ERK^*)cPLA_2 \xrightarrow{k_{cat}} ERK^* + cPLA_2^P,$ $k_{cat} = 14.0 \text{ s}^{-1}$	(Waas and Dalby, 2002; Waas et al., 2003)
Reac122	$ERK^* + (Ca^{2+})cPLA_2 \xrightarrow{k_f} (ERK^*)(Ca^{2+})cPLA_2,$ $k_f = 4.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Waas and Dalby, 2002; Waas et al., 2003)
Reac123	$(ERK^*)(Ca^{2+})cPLA_2 \xrightarrow{k_b} ERK^* + (Ca^{2+})cPLA_2,$ $k_b = 1.0 \text{ s}^{-1}$	(Waas and Dalby, 2002; Waas et al., 2003)
Reac124	$(ERK^*)(Ca^{2+})cPLA_2 \xrightarrow{k_{cat}} ERK^* + (Ca^{2+})cPLA_2^P,$ $k_{cat} = 14.0 \text{ s}^{-1}$	(Waas and Dalby, 2002; Waas et al., 2003)
Reac125	$ERK^* + (Ca^{2+})_2cPLA_2 \xrightarrow{k_f} (ERK^*)(Ca^{2+})_2cPLA_2,$	(Waas and Dalby,

	$k_f = 4.0 \text{ } \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	2002; Waas et al., 2003)
Reac126	$(ERK^*)(Ca^{2+})_2 cPLA_2 \xrightarrow{k_b} ERK^* + (Ca^{2+})_2 cPLA_2,$ $k_b = 1.0 \text{ s}^{-1}$	(Waas and Dalby, 2002; Waas et al., 2003)
Reac127	$(ERK^*)(Ca^{2+})_2 cPLA_2 \xrightarrow{k_{cat}} ERK^* + (Ca^{2+})_2 cPLA_2^P,$ $k_{cat} = 14.0 \text{ s}^{-1}$	(Waas and Dalby, 2002; Waas et al., 2003)
Reac128	$PP2A + cPLA_2^P \xrightarrow{k_f} (PP2A)cPLA_2^P,$ $k_f = 1.4 \text{ } \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac129	$(PP2A)cPLA_2^P \xrightarrow{k_b} PP2A + cPLA_2^P,$ $k_b = 1.5 \text{ s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac130	$(PP2A)cPLA_2^P \xrightarrow{k_{cat}} PP2A + cPLA_2,$ $k_{cat} = 2.5 \text{ s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac131	$PP2A + (Ca^{2+})cPLA_2^P \xrightarrow{k_f} (PP2A)(Ca^{2+})cPLA_2^P,$ $k_f = 1.4 \text{ } \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac132	$(PP2A)(Ca^{2+})cPLA_2^P \xrightarrow{k_b} PP2A + (Ca^{2+})cPLA_2^P,$ $k_b = 1.5 \text{ s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac133	$(PP2A)(Ca^{2+})cPLA_2^P \xrightarrow{k_{cat}} PP2A + (Ca^{2+})cPLA_2,$ $k_{cat} = 2.5 \text{ s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)

Reac134	$PP2A + (Ca^{2+})_2cPLA_2^P \xrightarrow{k_f} (PP2A)(Ca^{2+})_2cPLA_2^P,$ $k_f = 1.4 \text{ } \mu\text{mol}^{-1} \cdot \text{L.s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac135	$(PP2A)(Ca^{2+})_2cPLA_2^P \xrightarrow{k_b} PP2A + (Ca^{2+})_2cPLA_2^P,$ $k_b = 1.5 \text{ } \text{s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac136	$(PP2A)(Ca^{2+})_2cPLA_2^P \xrightarrow{k_{cat}} PP2A + (Ca^{2+})_2cPLA_2,$ $k_{cat} = 2.5 \text{ } \text{s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac137	$PP1 + cPLA_2^P \xrightarrow{k_f} (PP1)cPLA_2^P,$ $k_f = 1.4 \text{ } \mu\text{mol}^{-1} \cdot \text{L.s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac138	$(PP1)cPLA_2^P \xrightarrow{k_b} PP1 + cPLA_2^P,$ $k_b = 1.5 \text{ } \text{s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac139	$(PP1)cPLA_2^P \xrightarrow{k_{cat}} PP1 + cPLA_2,$ $k_{cat} = 2.5 \text{ } \text{s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac140	$PP1 + (Ca^{2+})cPLA_2^P \xrightarrow{k_f} (PP1)(Ca^{2+})cPLA_2^P,$ $k_f = 1.4 \text{ } \mu\text{mol}^{-1} \cdot \text{L.s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac141	$(PP1)(Ca^{2+})cPLA_2^P \xrightarrow{k_b} PP1 + (Ca^{2+})cPLA_2^P,$ $k_b = 1.5 \text{ } \text{s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992;

		Agostinis et al., 1996)
Reac142	$(PP1)(Ca^{2+})_2cPLA_2^P \xrightarrow{k_{cat}} PP1 + (Ca^{2+})_2cPLA_2,$ $k_{cat} = 2.5 \text{ s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac143	$PP1 + (Ca^{2+})_2cPLA_2^P \xrightarrow{k_f} (PP1)(Ca^{2+})_2cPLA_2^P,$ $k_f = 1.4 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac144	$(PP1)(Ca^{2+})_2cPLA_2^P \xrightarrow{k_b} PP1 + (Ca^{2+})_2cPLA_2^P,$ $k_b = 1.5 \text{ s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac145	$(PP1)(Ca^{2+})_2cPLA_2^P \xrightarrow{k_{cat}} PP1 + (Ca^{2+})_2cPLA_2^P,$ $k_{cat} = 2.5 \text{ s}^{-1}$	(Agostinis et al., 1987; Agostinis et al., 1992; Agostinis et al., 1996)
Reac146	$cPLA_2^P \xrightarrow{k_f} cPLA_2^{**},$ $k_f = 50.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Das et al., 2003; Stahelin et al., 2007)
Reac147	$cPLA_2^{**} \xrightarrow{k_b} cPLA_2^P,$ $k_b = 15.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Das et al., 2003; Stahelin et al., 2007)

Reac148	$cPLA_2^{**} \xrightarrow{k_f} (cPLA_2^{**})APC$, $k_f = 43.0 \text{ s}^{-1}$, pseudo-first order reaction of interaction between cPLA ₂ and its substrate	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac149	$(cPLA_2^{**})APC \xrightarrow{k_b} cPLA_2^{**}$, $k_b = 600.0 \text{ s}^{-1}$	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac150	$(cPLA_2^{**})APC \xrightarrow{k_{cat}} cPLA_2^{**} + AA$, $k_{cat} = 3600.0 \text{ s}^{-1}$	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac151	$(Ca^{2+})cPLA_2^P \xrightarrow{k_f} (Ca^{2+})cPLA_2^{**}$, $k_f = 50.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Das et al., 2003; Stahelin et al., 2007)
Reac152	$(Ca^{2+})cPLA_2^{**} \xrightarrow{k_b} (Ca^{2+})cPLA_2^P$, $k_b = 15.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Das et al., 2003; Stahelin et al., 2007)
Reac153	$(Ca^{2+})cPLA_2^{**} \xrightarrow{k_f} ((Ca^{2+})cPLA_2^{**})APC$, $k_f = 43.0 \text{ s}^{-1}$, pseudo-first order reaction of interaction between cPLA ₂ and its substrate	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac154	$((Ca^{2+})cPLA_2^{**})APC \xrightarrow{k_b} (Ca^{2+})cPLA_2^{**}$,	(Bayburt and Gelb,

	$k_b = 600.0 \text{ s}^{-1}$	1997; Berg et al., 2001; Tucker et al., 2009)
Reac155	$((Ca^{2+})_2cPLA_2^{**})APC \xrightarrow{k_{cat}} (Ca^{2+})_2cPLA_2^{**} + AA,$ $k_{cat} = 3600.0 \text{ s}^{-1}$	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac156	$(Ca^{2+})_2cPLA_2^P \xrightarrow{k_f} (Ca^{2+})_2cPLA_2^{**},$ $k_f = 300.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Das et al., 2003; Stahelin et al., 2007)
Reac157	$(Ca^{2+})_2cPLA_2^{**} \xrightarrow{k_b} (Ca^{2+})_2cPLA_2^P,$ $k_b = 15.0 \text{ s}^{-1}$	(Hixon et al., 1998; Das and Cho, 2002; Frazier et al., 2002; Das et al., 2003; Stahelin et al., 2007)
Reac158	$(Ca^{2+})_2cPLA_2^{**} \xrightarrow{k_f} ((Ca^{2+})_2cPLA_2^{**})APC,$ $k_f = 43.0 \text{ s}^{-1}$, pseudo-first order reaction of interaction between cPLA ₂ and its substrate	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac159	$((Ca^{2+})_2cPLA_2^{**})APC \xrightarrow{k_b} (Ca^{2+})_2cPLA_2^{**},$ $k_b = 600.0 \text{ s}^{-1}$	(Bayburt and Gelb, 1997; Berg et al., 2001; Tucker et al., 2009)
Reac160	$((Ca^{2+})_2cPLA_2^{**})APC \xrightarrow{k_{cat}} (Ca^{2+})_2cPLA_2^{**} + AA,$ $k_{cat} = 3600.0 \text{ s}^{-1}$	(Bayburt and Gelb, 1997; Berg et al., 2001)

		Tucker et al., 2009)
Reac161	$cPLA_2^P + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})cPLA_2^P$, $k_f = 1.93 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac162	$(Ca^{2+})cPLA_2^P \xrightarrow{k_b} cPLA_2^P + Ca^{2+}$, $k_b = 108 \text{ s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac163	$cPLA_2^{**} + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})cPLA_2^{**}$, $k_f = 1.93 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac164	$(Ca^{2+})cPLA_2^{**} \xrightarrow{k_b} cPLA_2^{**} + Ca^{2+}$, $k_b = 0.41 \text{ s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac165	$(Ca^{2+})cPLA_2^P + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2cPLA_2^P$, $k_f = 10.8 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke,

		2002)
Reac166	$(Ca^{2+})_2 cPLA_2^P \xrightarrow{k_b} (Ca^{2+})_2 cPLA_2^P + Ca^{2+}$, $k_b = 108 \text{ s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac167	$(Ca^{2+})cPLA_2^{**} + Ca^{2+} \xrightarrow{k_f} (Ca^{2+})_2 cPLA_2^{**}$, $k_f = 10.8 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac168	$(Ca^{2+})_2 cPLA_2^{**} \xrightarrow{k_b} (Ca^{2+})cPLA_2^{**} + Ca^{2+}$, $k_b = 2.5 \text{ s}^{-1}$	(Nalefski et al., 1997; Bittova et al., 1999; Nalefski et al., 2001; Nalefski and Falke, 2002)
Reac169	$AA \xrightarrow{k_{deg}} \emptyset$, $k_{deg} = 0.4 \text{ s}^{-1}$	(Bhalla and Iyengar, 1999; Tanaka et al., 2007)
Reac170	$(Ca^{2+})_3 PKC^* + AMPAR_{syn} \xrightarrow{k_f} ((Ca^{2+})_3 PKC^*)AMPAR_{syn}^f$, $k_f = 0.4 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Woodgett et al., 1986)
Reac171	$((Ca^{2+})_3 PKC^*)AMPAR_{syn} \xrightarrow{k_b} (Ca^{2+})_3 PKC^* + AMPAR_{syn}$, $k_b = 0.8 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac172	$((Ca^{2+})_3 PKC^*)AMPAR_{syn} \xrightarrow{k_{cat}} (Ca^{2+})_3 PKC^* + AMPAR_{syn}^P$, $k_{cat} = 0.3 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac173	$(AA)PKC^* + AMPAR_{syn} \xrightarrow{k_f} ((AA)PKC^*)AMPAR_{syn}$, $k_f = 0.4 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Woodgett et al., 1986)
Reac174	$((AA)PKC^*)AMPAR_{syn} \xrightarrow{k_b} (AA)PKC^* + AMPAR_{syn}$,	(Woodgett et al., 1986)

	$k_b = 0.8 \text{ s}^{-1}$	
Reac175	$((AA)PKC^*)AMPAR_{syn} \xrightarrow{k_{cat}} (AA)PKC^* + AMPAR_{syn}^P,$ $k_{cat} = 0.3 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac176	$(AA)(Ca^{2+})PKC^* + AMPAR_{syn} \xrightarrow{k_f} ((AA)(Ca^{2+})PKC^*)AMPAR_{syn},$ $k_f = 0.4 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Woodgett et al., 1986)
Reac177	$((AA)(Ca^{2+})PKC^*)AMPAR_{syn} \xrightarrow{k_b} (AA)(Ca^{2+})PKC^* + AMPAR_{syn},$ $k_b = 0.8 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac178	$((AA)(Ca^{2+})PKC^*)AMPAR_{syn} \xrightarrow{k_{cat}} (AA)(Ca^{2+})PKC^* + AMPAR_{syn}^P,$ $k_{cat} = 0.3 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac179	$(AA)(Ca^{2+})_3PKC^* + AMPAR_{syn} \xrightarrow{k_f} ((AA)(Ca^{2+})_3PKC^*)AMPAR_{syn},$ $k_f = 0.4 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Woodgett et al., 1986)
Reac180	$((AA)(Ca^{2+})_3PKC^*)AMPAR_{syn} \xrightarrow{k_b} (AA)(Ca^{2+})_3PKC^* + AMPAR_{syn},$ $k_b = 0.8 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac181	$((AA)(Ca^{2+})_3PKC^*)AMPAR_{syn} \xrightarrow{k_{cat}} (AA)(Ca^{2+})_3PKC^* + AMPAR_{syn}^P,$ $k_{cat} = 0.3 \text{ s}^{-1}$	(Woodgett et al., 1986)
Reac182	$PP2A + AMPAR_{syn}^P \xrightarrow{k_f} (PP2A)AMPAR_{syn}^P,$ $k_f = 0.6 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Donella Deana et al., 1990)
Reac183	$(PP2A)AMPAR_{syn}^P \xrightarrow{k_b} PP2A + AMPAR_{syn}^P,$ $k_b = 0.17 \text{ s}^{-1}$	(Donella Deana et al., 1990)
Reac184	$(PP2A)AMPAR_{syn}^P \xrightarrow{k_{cat}} PP2A + AMPAR_{syn},$ $k_{cat} = 0.25 \text{ s}^{-1}$	(Donella Deana et al., 1990)
Reac185	$GRIP + AMPAR_{syn} \xrightarrow{k_f} (GRIP)AMPAR_{syn},$ $k_f = 1.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Gianni et al., 2005; Gianni et al., 2006; Launey, 2007)
Reac186	$(GRIP)AMPAR_{syn} \xrightarrow{k_b} GRIP + AMPAR_{syn},$ $k_b = 7 \text{ s}^{-1}$	(Gianni et al., 2005; Gianni et al., 2006; Launey, 2007)
Reac187	$GRIP + AMPAR_{syn}^P \xrightarrow{k_f} (GRIP)AMPAR_{syn}^P,$ $k_f = 1.0 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Matsuda et al., 1999; Chung et al., 2003;

		Launey, 2007)
Reac188	$(GRIP)AMPAR_{syn}^P \xrightarrow{k_b} GRIP + AMPAR_{syn}^P,$ $k_b = 70 \text{ s}^{-1}$	(Matsuda et al., 1999; Chung et al., 2003; Launey, 2007)
Reac189	$AMPAR_{syn} \xrightarrow{k_f} AMPAR_{extra-syn},$ $k_f = 0.1 \text{ s}^{-1}$	(Borgdorff and Choquet, 2002; Bats et al., 2007)
Reac190	$AMPAR_{extra-syn} \xrightarrow{k_b} AMPAR_{syn},$ $k_b = 0.02 \text{ s}^{-1}$	(Borgdorff and Choquet, 2002; Bats et al., 2007)
Reac191	$AMPAR_{syn}^P \xrightarrow{k_f} AMPAR_{extra-syn}^P,$ $k_f = 0.1 \text{ s}^{-1}$	(Borgdorff and Choquet, 2002; Bats et al., 2007)
Reac192	$AMPAR_{extra-syn}^P \xrightarrow{k_b} AMPAR_{syn}^P,$ $k_b = 0.025 \text{ s}^{-1}$	(Borgdorff and Choquet, 2002; Bats et al., 2007)
Reac193	$PP2A + AMPAR_{extra-syn}^P \xrightarrow{k_f} (PP2A)AMPAR_{extra-syn}^P,$ $k_f = 0.6 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Donella Deana et al., 1990)
Reac194	$(PP2A)AMPAR_{extra-syn}^P \xrightarrow{k_b} PP2A + AMPAR_{extra-syn}^P,$ $k_b = 0.17 \text{ s}^{-1}$	(Donella Deana et al., 1990)
Reac195	$(PP2A)AMPAR_{extra-syn}^P \xrightarrow{k_{cat}} PP2A + AMPAR_{extra-syn},$ $k_{cat} = 0.25 \text{ s}^{-1}$	(Donella Deana et al., 1990)
Reac196	$AMPAR_{extra-syn} \xrightarrow{k_f} AMPAR_{dend},$ $k_f = 0.02 \text{ s}^{-1}$	(Borgdorff and Choquet, 2002; Bats et al., 2007)
Reac197	$AMPAR_{dend} \xrightarrow{k_b} AMPAR_{extra-syn},$ $k_b = 0.00025 \text{ s}^{-1}$	(Borgdorff and Choquet, 2002; Bats

		et al., 2007)
Reac198	$AMPAR_{extra-syn}^P \xrightarrow{k_f} AMPAR_{dend}^P,$ $k_f = 0.02 \text{ s}^{-1}$	(Borgdorff and Choquet, 2002; Bats et al., 2007)
Reac199	$AMPAR_{dend}^P \xrightarrow{k_b} AMPAR_{extra-syn}^P,$ $k_b = 0.00025 \text{ s}^{-1}$	(Borgdorff and Choquet, 2002; Bats et al., 2007)
Reac200	$PP2A + AMPAR_{dend}^P \xrightarrow{k_f} (PP2A)AMPAR_{dend}^P,$ $k_f = 0.6 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Donella Deana et al., 1990)
Reac201	$(PP2A)AMPAR_{dend}^P \xrightarrow{k_b} PP2A + AMPAR_{dend}^P,$ $k_b = 0.17 \text{ s}^{-1}$	(Donella Deana et al., 1990)
Reac202	$(PP2A)AMPAR_{dend}^P \xrightarrow{k_{cat}} PP2A + AMPAR_{dend},$ $k_{cat} = 0.25 \text{ s}^{-1}$	(Donella Deana et al., 1990)
Reac203	$AMPAR_{dend}^P \xrightarrow{k_f} AMPAR_{cyt}^P,$ $k_f = 0.003 \text{ s}^{-1}$	(Ehlers, 2000; Lin et al., 2000; Passafaro et al., 2001)
Reac204	$AMPAR_{cyt}^P \xrightarrow{k_b} AMPAR_{dend}^P,$ $k_b = 0.002 \text{ s}^{-1}$	(Ehlers, 2000; Passafaro et al., 2001)
Reac205	$PP2A + AMPAR_{cyt}^P \xrightarrow{k_f} (PP2A)AMPAR_{cyt}^P,$ $k_f = 0.6 \mu\text{mol}^{-1} \cdot \text{L} \cdot \text{s}^{-1}$	(Donella Deana et al., 1990)
Reac206	$(PP2A)AMPAR_{cyt}^P \xrightarrow{k_b} PP2A + AMPAR_{cyt}^P,$ $k_b = 0.17 \text{ s}^{-1}$	(Donella Deana et al., 1990)
Reac207	$(PP2A)AMPAR_{cyt}^P \xrightarrow{k_{cat}} PP2A + AMPAR_{cyt},$ $k_{cat} = 0.25 \text{ s}^{-1}$	(Donella Deana et al., 1990)

^a $1 \mu\text{mol} \cdot \text{L}^{-1}$ corresponds to ~ 49 molecules

^b Parenthesis indicates complex formation

^c The presence of * or ** indicates activated form of the species considered

^d The letter ^P indicates phosphorylation without activation

^e The term *memb* refers to membrane

^f The terms AMPAR_{syn}, AMPAR_{extra-syn}, AMPAR_{dend}, AMPAR_{cyt} refer, respectively, to synaptic AMPAR, extra-synaptic AMPAR in the spine, dendritic AMPAR and cytosolic AMPAR.

Table 1 References

- Agostinis P, Derua R, Sarno S, Goris J, Merlevede W (1992) Specificity of the polycation-stimulated (type-2A) and ATP,Mg-dependent (type-1) protein phosphatases toward substrates phosphorylated by P34cdc2 kinase. *Eur J Biochem* 205:241-248.
- Agostinis P, Goris J, Waelkens E, Pinna LA, Marchiori F, Merlevede W (1987) Dephosphorylation of phosphoproteins and synthetic phosphopeptides. Study of the specificity of the polycation-stimulated and MgATP-dependent phosphorylase phosphatases. *J Biol Chem* 262:1060-1064.
- Agostinis P, Donella-Deana A, Van Hoof C, Cesaro L, Brunati AM, Ruzzene M, Merlevede W, Pinna LA, Goris J (1996) A comparative study of the phosphotyrosyl phosphatase specificity of protein phosphatase type 2A and phosphotyrosyl phosphatase type 1B using phosphopeptides and the phosphoproteins p50/HS1, c-Fgr and Lyn. *Eur J Biochem* 236:548-557.
- Aksan I, Kurnaz ML (2003) A computer-based model for the regulation of mitogen activated protein kinase (MAPK) activation. *J Recept Signal Transduct Res* 23:197-209.
- Bahl R, Bradley KC, Thompson KJ, Swain RA, Rossie S, Meisel RL (2001) Localization of protein Ser/Thr phosphatase 5 in rat brain. *Brain Res Mol Brain Res* 90:101-109.
- Bats C, Groc L, Choquet D (2007) The interaction between Stargazin and PSD-95 regulates AMPA receptor surface trafficking. *Neuron* 53:719-734.
- Bayburt T, Gelb MH (1997) Interfacial catalysis by human 85 kDa cytosolic phospholipase A2 on anionic vesicles in the scooting mode. *Biochemistry* 36:3216-3231.
- Berg OG, Gelb MH, Tsai MD, Jain MK (2001) Interfacial enzymology: the secreted phospholipase A(2)-paradigm. *Chem Rev* 101:2613-2654.
- Bhalla US, Iyengar R (1999) Emergent properties of networks of biological signaling pathways. *Science* 283:381-387.
- Bittova L, Sumandea M, Cho W (1999) A structure-function study of the C2 domain of cytosolic phospholipase A2. Identification of essential calcium ligands and hydrophobic membrane binding residues. *J Biol Chem* 274:9665-9672.
- Bittova L, Stahelin RV, Cho W (2001) Roles of ionic residues of the C1 domain in protein kinase C-alpha activation and the origin of phosphatidylserine specificity. *J Biol Chem* 276:4218-4226.
- Borgdorff AJ, Choquet D (2002) Regulation of AMPA receptor lateral movements. *Nature* 417:649-653.
- Brinkmann T, Daumke O, Herbrand U, Kühlmann D, Stege P, Ahmadian MR, Wittinghofer A (2002) Rap-specific GTPase activating protein follows an alternative mechanism. *J Biol Chem* 277:12525-12531.

- Cheng D, Hoogenraad CC, Rush J, Ramm E, Schlager MA, Duong DM, Xu P, Wijayawardana SR, Hanfelt J, Nakagawa T, Sheng M, Peng J (2006) Relative and absolute quantification of postsynaptic density proteome isolated from rat forebrain and cerebellum. *Mol Cell Proteomics* 5:1158-1170.
- Chung HJ, Steinberg JP, Huganir RL, Linden DJ (2003) Requirement of AMPA receptor GluR2 phosphorylation for cerebellar long-term depression. *Science* 300:1751-1755.
- Das S, Cho W (2002) Roles of catalytic domain residues in interfacial binding and activation of group IV cytosolic phospholipase A2. *J Biol Chem* 277:23838-23846.
- Das S, Rafter JD, Kim KP, Gygi SP, Cho W (2003) Mechanism of group IVA cytosolic phospholipase A(2) activation by phosphorylation. *J Biol Chem* 278:41431-41442.
- Doi T, Kuroda S, Michikawa T, Kawato M (2005) Inositol 1,4,5-trisphosphate-dependent Ca²⁺ threshold dynamics detect spike timing in cerebellar Purkinje cells. *J Neurosci* 25:950-961.
- Donella Deana A, Mac Gowan CH, Cohen P, Marchiori F, Meyer HE, Pinna LA (1990) An investigation of the substrate specificity of protein phosphatase 2C using synthetic peptide substrates; comparison with protein phosphatase 2A. *Biochim Biophys Acta* 1051:199-202.
- Ehlers MD (2000) Reinsertion or degradation of AMPA receptors determined by activity-dependent endocytic sorting. *Neuron* 28:511-525.
- Frazier AA, Wisner MA, Malmberg NJ, Victor KG, Fanucci GE, Nalefski EA, Falke JJ, Cafiso DS (2002) Membrane orientation and position of the C2 domain from cPLA2 by site-directed spin labeling. *Biochemistry* 41:6282-6292.
- Fujioka A, Terai K, Itoh RE, Aoki K, Nakamura T, Kuroda S, Nishida E, Matsuda M (2006) Dynamics of the Ras/ERK MAPK cascade as monitored by fluorescent probes. *J Biol Chem* 281:8917-8926.
- Gianni S, Engström A, Larsson M, Calosci N, Malatesta F, Eklund L, Ngang CC, Travagliini-Allocatelli C, Jemth P (2005) The kinetics of PDZ domain-ligand interactions and implications for the binding mechanism. *J Biol Chem* 280:34805-34812.
- Gianni S, Walma T, Arcovito A, Calosci N, Bellelli A, Engström A, Travagliini-Allocatelli C, Brunori M, Jemth P, Vuister GW (2006) Demonstration of long-range interactions in a PDZ domain by NMR, kinetics, and protein engineering. *Structure* 14:1801-1809.
- Harris KM, Stevens JK (1988) Dendritic spines of rat cerebellar Purkinje cells: serial electron microscopy with reference to their biophysical characteristics. *J Neurosci* 8:4455-4469.
- Hixon MS, Ball A, Gelb MH (1998) Calcium-dependent and -independent interfacial binding and catalysis of cytosolic group IV phospholipase A2. *Biochemistry* 37:8516-8526.
- Huang CY, Ferrell JE (1996) Ultrasensitivity in the mitogen-activated protein kinase cascade. *Proc Natl Acad Sci U S A* 93:10078-10083.
- Keranen LM, Newton AC (1997) Ca²⁺ differentially regulates conventional protein kinase Cs' membrane interaction and activation. *J Biol Chem* 272:25959-25967.

- Kiyatkin A, Aksamitiene E, Markevich NI, Borisov NM, Hoek JB, Kholodenko BN (2006) Scaffolding protein Grb2-associated binder 1 sustains epidermal growth factor-induced mitogenic and survival signaling by multiple positive feedback loops. *J Biol Chem* 281:19925-19938.
- Kohout SC, Corbalán-García S, Torrecillas A, Goméz-Fernández JC, Falke JJ (2002) C2 domains of protein kinase C isoforms alpha, beta, and gamma: activation parameters and calcium stoichiometries of the membrane-bound state. *Biochemistry* 41:11411-11424.
- Launey T (2007) A computational approach to the study of AMPA receptor clustering at Purkinje cell synapses. *Arch Ital Biol* 145:299-310.
- Lenzen C, Cool RH, Prinz H, Kuhlmann J, Wittinghofer A (1998) Kinetic analysis by fluorescence of the interaction between Ras and the catalytic domain of the guanine nucleotide exchange factor Cdc25Mm. *Biochemistry* 37:7420-7430.
- Lin JW, Ju W, Foster K, Lee SH, Ahmadian G, Wyszynski M, Wang YT, Sheng M (2000) Distinct molecular mechanisms and divergent endocytotic pathways of AMPA receptor internalization. *Nat Neurosci* 3:1282-1290.
- Markevich NI, Hoek JB, Kholodenko BN (2004) Signaling switches and bistability arising from multisite phosphorylation in protein kinase cascades. *J Cell Biol* 164:353-359.
- Masugi-Tokita M, Tarusawa E, Watanabe M, Molnár E, Fujimoto K, Shigemoto R (2007) Number and density of AMPA receptors in individual synapses in the rat cerebellum as revealed by SDS-digested freeze-fracture replica labeling. *J Neurosci* 27:2135-2144.
- Matsuda S, Mikawa S, Hirai H (1999) Phosphorylation of serine-880 in GluR2 by protein kinase C prevents its C terminus from binding with glutamate receptor-interacting protein. *J Neurochem* 73:1765-1768.
- Momiyama A, Silver RA, Hausser M, Notomi T, Wu Y, Shigemoto R, Cull-Candy SG (2003) The density of AMPA receptors activated by a transmitter quantum at the climbing fibre-Purkinje cell synapse in immature rats. *J Physiol* 549:75-92.
- Nalefski EA, Falke JJ (2002) Cation charge and size selectivity of the C2 domain of cytosolic phospholipase A(2). *Biochemistry* 41:1109-1122.
- Nalefski EA, Slazas MM, Falke JJ (1997) Ca²⁺-signaling cycle of a membrane-docking C2 domain. *Biochemistry* 36:12011-12018.
- Nalefski EA, Wisner MA, Chen JZ, Sprang SR, Fukuda M, Mikoshiba K, Falke JJ (2001) C2 domains from different Ca²⁺ signaling pathways display functional and mechanistic diversity. *Biochemistry* 40:3089-3100.
- Newton AC (2001) Protein kinase C: structural and spatial regulation by phosphorylation, cofactors, and macromolecular interactions. *Chem Rev* 101:2353-2364.
- Newton AC (2009) Lipid activation of protein kinases. *J Lipid Res* 50 Suppl:S266-271.
- O'Flaherty JT, Chadwell BA, Kearns MW, Sergeant S, Daniel LW (2001) Protein kinases C translocation responses to low concentrations of arachidonic acid. *J Biol Chem* 276:24743-24750.
- Passafaro M, Piëch V, Sheng M (2001) Subunit-specific temporal and spatial patterns of AMPA receptor exocytosis in hippocampal neurons. *Nat Neurosci* 4:917-926.

- Pearson G, Robinson F, Beers Gibson T, Xu BE, Karandikar M, Berman K, Cobb MH (2001) Mitogen-activated protein (MAP) kinase pathways: regulation and physiological functions. *Endocr Rev* 22:153-183.
- Rossie S, Jayachandran H, Meisel RL (2006) Cellular co-localization of protein phosphatase 5 and glucocorticoid receptors in rat brain. *Brain Res* 1111:1-11.
- Schaechter JD, Benowitz LI (1993) Activation of protein kinase C by arachidonic acid selectively enhances the phosphorylation of GAP-43 in nerve terminal membranes. *J Neurosci* 13:4361-4371.
- Schmidt H, Stiefel KM, Racay P, Schwaller B, Eilers J (2003) Mutational analysis of dendritic Ca²⁺ kinetics in rodent Purkinje cells: role of parvalbumin and calbindin D28k. *J Physiol* 551:13-32.
- Shinomura T, Asaoka Y, Oka M, Yoshida K, Nishizuka Y (1991) Synergistic action of diacylglycerol and unsaturated fatty acid for protein kinase C activation: its possible implications. *Proc Natl Acad Sci U S A* 88:5149-5153.
- Stahelin RV, Subramanian P, Vora M, Cho W, Chalfant CE (2007) Ceramide-1-phosphate binds group IVA cytosolic phospholipase a2 via a novel site in the C2 domain. *J Biol Chem* 282:20467-20474.
- Tanaka K, Khiroug L, Santamaria F, Doi T, Ogasawara H, Ellis-Davies GC, Kawato M, Augustine GJ (2007) Ca²⁺ requirements for cerebellar long-term synaptic depression: role for a postsynaptic leaky integrator. *Neuron* 54:787-800.
- Torrecillas A, Laynez J, Menéndez M, Corbalán-García S, Gómez-Fernández JC (2004) Calorimetric study of the interaction of the C2 domains of classical protein kinase C isoenzymes with Ca²⁺ and phospholipids. *Biochemistry* 43:11727-11739.
- Tucker DE, Ghosh M, Ghomashchi F, Loper R, Suram S, John BS, Girotti M, Bollinger JG, Gelb MH, Leslie CC (2009) Role of phosphorylation and basic residues in the catalytic domain of cytosolic phospholipase A2alpha in regulating interfacial kinetics and binding and cellular function. *J Biol Chem* 284:9596-9611.
- Waas WF, Dalby KN (2002) Transient protein-protein interactions and a random-ordered kinetic mechanism for the phosphorylation of a transcription factor by extracellular-regulated protein kinase 2. *J Biol Chem* 277:12532-12540.
- Waas WF, Rainey MA, Szafranska AE, Dalby KN (2003) Two rate-limiting steps in the kinetic mechanism of the serine/threonine specific protein kinase ERK2: a case of fast phosphorylation followed by fast product release. *Biochemistry* 42:12273-12286.
- Woodgett JR, Gould KL, Hunter T (1986) Substrate specificity of protein kinase C. Use of synthetic peptides corresponding to physiological sites as probes for substrate recognition requirements. *Eur J Biochem* 161:177-184.
- Zhao Y, Zhang ZY (2001) The mechanism of dephosphorylation of extracellular signal-regulated kinase 2 by mitogen-activated protein kinase phosphatase 3. *J Biol Chem* 276:32382-32391.
- Zhou B, Wang ZX, Zhao Y, Brautigan DL, Zhang ZY (2002) The specificity of extracellular signal-regulated kinase 2 dephosphorylation by protein phosphatases. *J Biol Chem* 277:31818-31825.

Table 2 Sensitivity analysis of the biochemical population size of the components of the positive feedback loop used in the model. Parameters (n_{Hill} , $K_{1/2}$, LTD_{max}) were estimated with 95% confidence interval by nonlinear least square regression to Equation 1.

Specie	n_{Hill}	$K_{1/2}$ ($\mu\text{mol.L}^{-1}$)	LTD_{max} (%)
Optimal model	5.72	1.50	32.80
PKC +10%	5.29 (7.52 %) ^a	1.36 (9.34 %)	33.36 (1.70 %)
PKC -10%	5.28 (7.69 %)	1.64 (9.34 %)	29.62 (9.65 %)
Raf-act +10%	5.84 (2.09)	1.43 (4.67 %)	31.99 (2.47 %)
Raf-act -10%	5.69 (0.52 %)	1.56 (4.00 %)	30.95 (6.04 %)
Raf +10%	5.59 (2.27 %)	1.39 (7.34 %)	33.83 (3.14 %)
Raf -10%	5.79 (1.22 %)	1.64 (9.34 %)	28.28 (13.78 %)
MEK +10%	5.46 (4.55 %)	1.40 (6.67 %)	32.61 (0.58 %)
MEK -10%	6.23 (8.91 %)	1.58 (5.34 %)	30.17 (8.02 %)
ERK +10%	5.99 (4.72 %)	1.44 (4.00 %)	31.88 (2.80 %)
ERK -10%	5.56 (2.79 %)	1.55 (3.34 %)	30.09 (8.26 %)
cPLA ₂ +10%	5.41 (5.41 %)	1.44 (4.00 %)	31.91 (2.71 %)
cPLA ₂ -10%	6.04 (5.59 %)	1.59 (6.00 %)	30.16 (8.05 %)
PP1 +10%	5.52 (3.49 %)	1.48 (1.34 %)	31.94 (2.62 %)
PP1 -10%	5.61 (1.92 %)	1.51 (0.67 %)	31.28 (4.63 %)
PP5 +10%	6.29 (9.96 %)	1.55 (3.34 %)	29.38 (10.43 %)
PP5 -10%	5.47 (4.37 %)	1.35 (10.0 %)	31.96 (2.56 %)
PP2A +10%	6.50 (13.63 %)	1.65 (10.0 %)	24.51 (25.27 %)
PP2A-10%	6.04 (5.59 %)	1.36 (9.34 %)	35.22 (7.37 %)
MKP +10%	6.07 (6.11 %)	1.52 (1.34 %)	28.12 (14.26 %)
MKP -10%	5.79 (1.22 %)	1.41 (6.00 %)	32.15 (1.98 %)

^aNumber in parenthesis show the percentage of the deviation in comparison with the mean response calculated for 156 runs of the optimal model

Table 3 Sensitivity analysis of the unknown rate constants of the reactions of the positive feedback loop used in the model. Parameters (n_{Hill} , $K_{1/2}$, LTD_{max}) were estimated with 95% confidence interval by nonlinear least square regression to Equation 1.

Rate constant identity	n_{Hill}	$K_{1/2}$ ($\mu\text{mol.L}^{-1}$)	LTD_{max} (%)
Optimal model	5.72	1.50	32.80
Rate constant 59, 62, 65 and 68 ^{a,b} +10%	5.50 (3.85 %) ^c	1.40 (6.67 %)	31.93 (2.65 %)
Rate constant 59, 62, 65 and 68 ^a - 10%	6.07 (6.12 %)	1.44 (4.00 %)	29.19 (11.00 %)
Rate constant 60, 63, 66 and 69 +10%	5.98 (4.55 %)	1.50 (0.00 %)	30.48 (7.07 %)
Rate constant 60, 63, 66 and 69 - 10%	6.26 (9.44 %)	1.46 (2.67 %)	30.97 (5.57 %)
Rate constant 61, 64, 67 and 70 +10%	5.75 (0.52 %)	1.48 (1.34 %)	31.94 (2.62 %)
Rate constant 61, 64, 67 and 70 - 10%	5.98 (4.55 %)	1.46 (2.67 %)	30.38 (7.37 %)
Rate constant 71 +10%	6.06 (5.94 %)	1.47 (2.00 %)	30.05 (8.38 %)
Rate constant 71 -10%	5.44 (4.89 %)	1.53 (2.00 %)	32.10 (2.13 %)
Rate constant 72 +10%	6.08 (6.29 %)	1.47 (2.00 %)	32.52 (0.85 %)
Rate constant 72 -10%	6.29 (9.96 %)	1.62 (8.00 %)	30.33 (7.53%)
Rate constant 73 +10%	6.18 (8.04 %)	1.50 (0.00 %)	29.84 (9.02 %)
Rate constant 73 -10%	6.21 (8.57 %)	1.49 (0.67 %)	31.12 (5.12 %)
Rate constant 74 +10%	6.03 (5.42 %)	1.44 (4.00 %)	31.53 (3.87 %)
Rate constant 74 -10%	6.16 (7.69 %)	1.49 (0.67 %)	29.15 (11.12 %)
Rate constant 75 +10%	5.37 (6.12 %)	1.53 (2.00 %)	29.56 (9.87 %)
Rate constant 75 -10%	6.24 (9.09 %)	1.45 (3.34 %)	32.94 (0.42 %)
Rate constant 76	6.22 (8.74 %)	1.41 (6.00 %)	32.02 (2.38 %)

+10%			
Rate constant 76	6.60 (15.38 %)	1.55 (3.34 %)	28.91 (11.85 %)
-10%			
Rate constant 77	5.53 (3.32 %)	1.52 (1.34 %)	29.71 (9.42 %)
+10%			
Rate constant 77	5.61 (1.92 %)	1.51 (0.67 %)	31.56 (3.78 %)
-10%			
Rate constant 84 and 87 +10% ^c	6.57 (14.86 %)	1.59 (6.00 %)	26.80 (18.29 %)
Rate constant 84 and 87 -10%	6.29 (9.96 %)	1.35 (10.00 %)	33.09 (0.88 %)
Rate constant 85 and 88 +10%	5.51 (3.67 %)	1.43 (4.67 %)	33.81 (3.07 %)
Rate constant 85 and 88 -10%	6.04 (5.59 %)	1.59 (6.00 %)	28.31 (13.69 %)
Rate constant 86 and 89 +10%	6.27 (9.61 %)	1.57 (4.67 %)	27.33 (16.68 %)
Rate constant 86 and 89 -10%	5.27 (7.86 %)	1.39 (7.34 %)	33.18 (1.16 %)
Rate constant 119, 122 and 125 +10%	5.91 (3.32 %)	1.54 (2.67 %)	31.47 (4.05 %)
Rate constant 119, 122 and 125 -10%	6.22 (8.74 %)	1.51 (0.67 %)	31.09 (5.21 %)
Rate constant 120, 123 and 126 +10%	6.19 (8.22 %)	1.47 (2.00 %)	30.67 (6.49 %)
Rate constant 120, 123 and 126 -10%	6.27 (9.61 %)	1.44 (4.00 %)	30.31 (7.59 %)
Rate constant 121, 124 and 127 +10%	6.28 (9.79 %)	1.52 (1.34 %)	30.69 (6.43 %)
Rate constant 121, 124 and 127 -10%	6.11 (6.82 %)	1.43 (4.67 %)	30.97 (5.58 %)
Rate constant 128, 131 and 134 +10%	6.19 (8.22 %)	1.51 (0.67 %)	30.42 (7.26 %)
Rate constant 128, 131 and 134 -10%	5.55 (2.97 %)	1.49 (0.67 %)	31.34 (4.45 %)
Rate constant 129, 132 and 135 +10%	5.69 (0.52 %)	1.48 (1.34 %)	31.52 (3.90 %)

Rate constant 129, 132 and 135 -10%	6.09 (6.47 %)	1.46 (2.67 %)	30.77 (6.19 %)
Rate constant 130, 133 and 136 +10%	5.56 (2.80 %)	1.47 (2.00 %)	30.14 (8.11 %)
Rate constant 130, 133 and 136 -10%	5.44 (5.39 %)	1.52 (1.34 %)	31.15 (5.03 %)
Rate constant 137, 140 and 143 +10%	6.27 (9.61 %)	1.49 (0.67 %)	30.36 (7.44 %)
Rate constant 137, 140 and 143 -10%	6.48 (13.28 %)	1.46 (2.67 %)	29.89 (8.87 %)
Rate constant 138, 141 and 144 +10%	6.14 (7.34 %)	1.43 (4.67 %)	30.86 (5.91 %)
Rate constant 138, 141 and 144 -10%	6.54 (14.33 %)	1.54 (2.67 %)	31.39 (4.29 %)
Rate constant 139, 142 and 145 +10%	5.24 (8.39 %)	1.48 (1.34 %)	31.91 (2.71 %)
Rate constant 139, 142 and 145 -10%	6.20 (8.39 %)	1.54 (2.67 %)	31.13 (5.09 %)
Rate constant 170, 173, 176 and 179 +10%	5.69 (0.52 %)	1.46 (2.67 %)	31.33 (4.48 %)
Rate constant 170, 173, 176 and 179 -10%	6.53 (14.16 %)	1.43 (4.67 %)	29.74 (9.32 %)
Rate constant 171, 174, 177 and 180 +10%	5.62 (1.75 %)	1.41 (6.00 %)	30.35 (7.47 %)
Rate constant 171, 174, 177 and 180 -10%	5.99 (4.72 %)	1.52 (1.34 %)	31.28 (4.63 %)
Rate constant 172, 175, 178 and 181 +10%	5.50 (3.85 %)	1.44 (4.00 %)	31.71 (3.32 %)
Rate constant 172, 175, 178 and 181 -10%	6.28 (9.79 %)	1.59 (6.00 %)	28.87 (11.98 %)
Rate constant 182,	6.02 (5.24 %)	1.47 (2.00 %)	29.69 (9.48 %)

193, 200 and 205 +10%			
Rate constant 182, 193, 200 and 205 -10%	6.28 (9.79 %)	1.46 (2.67 %)	31.36 (4.39 %)
Rate constant 183, 194, 201 and 206 +10%	6.13 (7.17 %)	1.49 (0.67 %)	30.65 (6.55 %)
Rate constant 183, 194, 201 and 206 -10%	6.27 (9.62 %)	1.53 (2.00 %)	30.32 (7.56 %)
Rate constant 184, 195, 202 and 207 +10%	5.20 (9.09 %)	1.54 (2.67 %)	30.72 (6.34 %)
Rate constant 184, 195, 202 and 207 -10%	5.34 (6.64 %)	1.44 (4.00 %)	31.56 (3.78 %)

^a The identities (numbers) of the rate constants in column 1 correspond to the same identities of the reactions (reac) listed in Supplementary Table I.

^b The parameters of analogous reactions were analyzed simultaneously.

^c Number in parenthesis show the percentage of the deviation in comparison with the mean responses of the optimal model.